### **ORIGINAL ARTICLE**

## MOLECULAR ECOLOGY WILEY

# Genomic differentiation and local adaptation on a microgeographic scale in a resident songbird

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#### Abstract

Elucidating forces capable of driving species diversification in the face of gene flow remains a key goal in evolutionary biology. Song sparrows, Melospiza melodia, occur as 25 subspecies in diverse habitats across North America, are among the continent's most widespread vertebrate species, and are exemplary of many highly variable species for which the conservation of locally adapted populations may be critical to their range-wide persistence. We focus here on six morphologically distinct subspecies resident in the San Francisco Bay region, including three salt-marsh endemics and three residents in upland and riparian habitats adjacent to the Bay. We used reducedrepresentation sequencing to generate 2,773 SNPs to explore genetic differentiation, spatial population structure, and demographic history. Clustering separated individuals from each of the six subspecies, indicating subtle differentiation at microgeographic scales. Evidence of limited gene flow and low nucleotide diversity across all six subspecies further supports a hypothesis of isolation among locally adapted populations. We suggest that natural selection for genotypes adapted to salt marsh environments and changes in demography over the past century have acted in concert to drive the patterns of diversification reported here. Our results offer evidence of microgeographic specialization in a highly polytypic bird species long discussed as a model of sympatric speciation and rapid adaptation, and they support the hypothesis that conserving locally adapted populations may be critical to the range-wide persistence of similarly highly variable species.

#### KEYWORDS

conservation units, demographic history, genetic structure, local adaptation, song sparrows

## **1** | INTRODUCTION

While geographic isolation is viewed as a predominant driver of divergence among populations, a substantial and growing literature suggests that evolutionary diversification can also arise via ecological selection across spatially heterogeneous environments at fine spatial scales (e.g., Kavanagh et al., 2010; Krueger-Hadfield et al., 2013; Langin et al., 2015; Richardson & Urban, 2013). Theory suggests that adaptive divergence at microgeographic scales (i.e., within the potential dispersal radius of an organism) can occur when natural selection overpowers the homogenizing effects of gene flow (Hendry et al., 2001; Richardson et al., 2014; Wright, 1969). Despite supporting theory (Bolnick & Fitzpatrick, 2007; Richardson et al., 2014), empirical studies of adaptation at microgeographic scales remain scarce, perhaps because sampling often occurs at scales exceeding the dispersal capabilities of the organisms under study (Richardson et al., 2014). Nevertheless, a number of classic examples of fine-scaled structuring in plant, insect, -WII FY-MOLECULAR ECOLOGY

and fish populations have appeared (Antonovics, 2006; Hendry et al., 2002; Nosil & Crespi, 2004). As a consequence, an emerging consensus suggests that local adaptation at microgeographic scales may be more common than once thought. This conclusion challenges the idea that population divergence requires periods of geographic isolation and supports the hypothesis that the conservation of locally adapted populations may be crucial to ensuring the persistence of species across their range (e.g., Aitken & Whitlock, 2013; Ho & Zhang, 2018; Reid et al., 2018).

Whereas we expect adaptive divergence to be greater when gene flow and genetic drift between populations are low (Charmantier et al., 2016), there is a growing appreciation for the complexity that arises when gene flow and selection are acting in concert. Gene flow can result in increased variation and genetic novelty, thus promoting divergence and adaptive potential (Garnett et al., 2007) or it can have an opposite, homogenizing effect that prevents divergence (Slatkin, 1985). Elucidating such processes thus requires further empirical research on species displaying evident diversification at fine spatial scales. Given that some of the best examples of microgeographic adaptation involve mainly sedentary organisms such as plants (Steiner & Berrang, 1990; Antonovics, 2006; Hendrick et al., 2016), we aimed to investigate such processes in a more mobile organism, wherein the signal of microgeographic adaptation may be less evident and underappreciated as a factor affecting the rate of diversification under gene flow.

Here we leverage a song sparrow "subspecies complex" resident in the San Francisco Bay region of California, USA (referred to as the Bay or bay region throughout this manuscript), to evaluate genomic differentiation at fine spatial scales ( $70 \times 100$  km) and spanning multiple fresh-to-salt water habitat gradients. At the broadest scale, the song sparrow (Melospiza melodia) is one of North America's most abundant, adaptable, and widespread bird species. However, at finer scales, song sparrows display differences in song, size, and plumage coloration that imply a history of local adaptation, particularly where birds reside on or near territories year-round (Arcese et al., 2002; Pruett & Winker, 2005). Despite being found in close geographic proximity, five recognized and phenotypically distinct subspecies of song sparrow reside year-round in the San Francisco Bay region (Figure 1a; Grinnell & Miller, 1944). M. m. samuelis (Samuels Song Sparrow in the marshes of the San Pablo Bay), M. m. maxillaris (Suisun Song Sparrow in the marshes of Suisun Bay), and M. m. pusillula (Alameda Song Sparrow in the marshes of South San Francisco Bay) are endemic subspecies found only in the saline marshes of their three respective sub-bays. M. m. gouldii (Marin Song Sparrow) occupies the surrounding upland regions along the coast and surrounding the Bay, and M. m. heermanni (Heermann's Song Sparrow) occupies riparian zones east of the Bay (Chan & Arcese, 2002). A sixth putative subspecies occupying upland habitat around the Bay, M. m. santaecrucis, was described by Grinnell (1901) and is thought to be the result of hybridization between M. m. gouldii and M. m. heermanni



**FIGURE 1** Map of the study site with illustrations of song sparrow subspecies to demonstrate morphological variation (a) and principal component analyses (PCA; b). The map of the San Francisco Bay shows the 12 sampling sites and the respective subspecies sampled (size of points indicates the number of individuals sampled at each site). *M. m. pusillula, maxillaris,* and *samuelis* are the three subspecies endemic to the salt marsh (blue colours), and *gouldii, heermanni,* and *santaecrucis* are the upland subspecies (red colours). PC1 demonstrates strong clustering of *pusillula,* PC2 explains more of the divergence in the upland subspecies, *gouldii* and *heermanni,* and PC3 explains variation between saltmarsh and upland populations. Song sparrow illustrations by Jillian Ditner (2019)

Subspecies	Locality	County, State	# of Individuals	Н <sub>о</sub>	π
M. m. gouldii	Mark's Marsh, Tomales Bay, Audubon Canyon Ranch (MM)	Marin County, California	24	0.1394	0.00135
	Tennessee Valley (TV)	Marin County, California	1	0.2157	0.002
M. m. heermanni	The Nature Conservancy Cosumnes River Preserve (CO)	Sacramento County, California	25	0.1081	0.001
M. m. santaecrucis	Los Gatos Creek County Park (LG)	Santa Clara County, California	18	0.1084	0.0011
	San Pedro Valley County Park (SP)	San Mateo County, California	7	0.1201	0.0013
M. m. maxillaris	Goodyear Slough Unit, Grizzly Island Wildlife Area (GS)	Solano County, California	14	0.1097	0.0011
	Rush Ranch Open Space, Solano County Farmlands and Open Space Foundation (RR)	Solano County, California	15	0.1102	0.0012
M. m. pusillula	Dumbarton Marsh, Don Edwards San Francisco Bay National Wildlife Refuge (DM)	Alameda County, California	14	0.1030	0.0011
	Palo Alto Baylands Nature Preserve (PB)	San Mateo County, California	15	0.1030	0.0011
M. m. samuelis	Petaluma River Mouth, California Department of Fish and Game (PM)	Sonoma County, California	6	0.1123	0.0012
	Sonoma Creek, San Pablo Bay National Wildlife Refuge (SC)	Solano County, California	19	0.1071	0.0011
	China Camp State Park (CC)	Marin Co, California	2	0.1443	0.0015

**TABLE 1** Sampling information for six subspecies of song sparrow (*Melospiza melodia*). For each subspecies, we report sampling location, sample size, observed heterozygosity ( $H_{a}$ ) and nucleotide diversity ( $\pi$ )

(Patten & Pruett, 2009). In 2008, four of these six subspecies were classified by the State of California as "Species of Special Concern:" *pusillula, maxillaris, samuelis, and heermanni* (formerly *M. m. mailliardi;* Northern Central Valley population). The further loss of marsh habitat represents a severe threat to the persistence of these endemic subspecies (Sauer et al., 2014) by raising the likelihood of extinction in a potentially unique group of song sparrows adapted to saline conditions.

The diversification of song sparrows in this region has received over a century of investigation into their morphological, genetic, and behavioural divergence (Chan & Arcese, 2002, 2003; Grinnell, 1913; Huxley, 1942; Johnston, 1956a, 1956b; Marshall, 1948a,1948b; Mayr, 1963; Miller, 1947, 1956). Results to date have revealed geographic structure in plumage and morphology, and blood type, but less evidence in behavioral or life history traits (Chan & Arcese, 2002, 2003; Ferrell, 1966; Johnston, 1956a, 1956b; Mulligan, 1963). Several mechanisms have been proposed to explain the diversification of song sparrows in San Francisco Bay, including habitat selection (Grinnell, 1913), habitat selection coupled with isolation (Marshall, 1948b), drift among populations of small demographic size (Ferrell, 1966; Miller, 1947), geographic isolation in different arms of the San Francisco Bay (Mayr, 1942), strong selection favouring phenotypic variation despite ongoing gene flow (Aldrich, 1984; Zink & Dittmann, 1993), and phenotypic plasticity in the absence of genetic variation (Smith, 1998; Zink & Dittmann, 1993). In contrast, prior studies of genetic variation in microsatellite (Chan & Arcese, 2002)

and mitochondrial DNA (Fry & Zink, 1998; Zink & Dittmann, 1993) suggest that divergence among subspecies in the bay region is low, despite representing diagnosable phenotypes.

The observed morphological variation among San Francisco Bay song sparrows is consistent with well-documented patterns of phenotypic convergence observed in tidal marsh sparrows in traits with putative adaptive functions to tidal environments (e.g., Basham et al., 1987; Greenberg & Olsen, 2010; Grenier & Greenberg, 2006; Luttrell et al., 2014; Tattersall et al., 2017; Walsh et al., 2019a). For example, maxillaris has a 40% greater bill depth-a hypothesized mechanism for facilitating evaporative heat loss in freshwater-limited and exposed environment (Greenberg et al., 2012; Tattersall et al., 2017)-than its upland counterpart gouldii, and experimental studies suggest significant salt tolerance in pusillula, whereas individual gouldii were unable to maintain bodyweight when consuming saline solutions (Basham et al., 1987). Taken together, these patterns are consistent with a hypothesis of phenotypic differentiation driven by strong selection on functional loci that influence traits intimately linked to individual performance in a local environment, and which reflect the outcome of adaptative evolution at microgeographic scales in the absence of differentiation at neutral loci.

To test this hypothesis, we elucidated further the microgeographic patterns of genetic structure in San Francisco Bay song sparrows by replicating the sampling design of Chan and Arcese (2002) to assess microsatellite diversification in the bay region using updated genomic methods and high-resolution sequencing. Specifically, our objectives 4 WILFY-MOLECULAR ECOLOGY

were to: (a) characterize patterns of song sparrow diversification in the San Francisco Bay; (b) employ our genomic markers to delineate population boundaries at a fine spatial scale; and (c) combine population genetic and demographic analyses with outlier scans for loci under divergent selection to estimate the contributions of previously proposed mechanisms underlying observed patterns of divergence in the region. To do so, we revisit existing hypotheses on the divergence of song sparrows in this region to evaluate the potential roles of genetic drift, geographic isolation, and local adaptation in divergence, and to speculate on the roles that local adaptation to tidal marsh environments may play in structuring populations. We conclude by discussing our results in the context of evolutionary theory and provide information to aid in the management of endemic song sparrows in the bay region based on degree of evolutionary divergence.

#### MATERIALS AND METHODS 2

#### 2.1 | Study system and sampling

All birds were sampled during the breeding season (March to May) by YLC and PA (see Chan & Arcese, 2002 for additional details). Blood was collected in heparinized capillary tubes and transferred to 1 ml of 1X lysis buffer and stored at 4°C. Individuals were sampled in 1999 from both tidal salt marshes and the upland riparian areas in the bay region (Table 1; Figure 1a). In total, we obtained samples from 160 individuals from 12 different populations representing all six of the putative subspecies in the region (Table S1). All blood samples used in this study are currently housed at the Cornell University Museum of Vertebrates.

#### 2.2 Molecular methods

Genomic DNA was extracted from blood samples using Qiagen DNeasy blood and tissue extraction kits (Qiagen) following the manufacturer's protocol. We used double-digest restriction siteassociated DNA sequencing (ddRAD) according to the protocol of Peterson et al. (2012) with modifications following Thrasher et al. (2017) to construct libraries and generate genomic data for single nucleotide polymorphism (SNP) discovery. In brief, we used ~200 to 500 ng of DNA for each individual (concentrations ranged from 8 to 57 ng/µl); concentrations were determined using the Qubit fluorometer and dsDNA broad range assay kit (ThermoFisher Scientific, Q32853, Life Technologies). DNA was digested with SbfI and Mspl restriction enzymes (New England Biolabs) and ligated to one of 20 P1 adaptors and a P2 adaptor using T4 DNA ligase (New England Biolabs). Samples with similar DNA concentrations were then separately pooled into a total of eight index groups. Samples were purified with MagNA beads prepared according to the protocol of Rohland and Reich (2012) to remove the enzymes and small DNA fragments. Fragments were size selected between 400 and 700 bp using the Blue Pippin (Sage Science) to ensure the same loci were recovered in all index groups. We performed low-cycle PCR with Phusion High-Fidelity DNA Polymerase (New England Biolabs) for each index group, purified with SPRI beads to eliminate small fragments, and then visualized the product on 1% agarose gel and fragment Bioanalyser (Agilent Technologies). The index groups were combined and sequenced on one Illumina HiSeq 2500 lane (read length 100 bp, single end) using the rapid run mode at the Cornell University Biotechnology Resource Center.

#### Data processing and SNP calling 2.3

Sequence quality was assessed using FASTQC version 0.11.8. (www. bioinformatics.babraham.ac.uk/projects/fastqc). We removed three samples due to poor quality. The remaining 157 individuals were filtered for quality using the FAST-X Toolkit (http://hannonlab.cshl.edu/ fastx\_toolkit), removing sequences with Phred quality scores below 10 and sequences with more than 5% of bases with Phred guality scores below 20. We demultiplexed sequences using the command "process\_radtags" in STACKS version 1.48 (Catchen et al., 2011) and additionally filtered samples to only retain reads that passed the Illumina chastity filter, contained an intact SbfI RAD site, contained a unique sample barcode, and did not contain Illumina indexing adapters. To account for differences in length, the remaining filtered and demultiplexed reads were trimmed to 94 base pairs at the 3' end using FAST-X TRIMMER (FAST-X Toolkit).

Sequences were aligned to a song sparrow (Melospiza melodia) reference genome (Feng et al., in review) using BOWTIE2 version 2.3. Mapped reads were then analysed using the ref\_map.pl pipeline in STACKS. We allowed five mismatches between sample loci and required a minimum of ten identical raw reads to make a stack. We ran the Populations module in STACKS for one population (-p) and required that a locus be present in a minimum of 80% of individuals to be processed (-r). In addition to obtaining all SNPs per locus (10,270), we created a subset of SNPs that included only the first SNP per stack (2,773). To avoid bias associated with physical linkage (O'Leary et al., 2018), we used the reduced, unlinked data set with 2,773 SNPs for all analyses, unless otherwise stated.

We used the "relatedness" function within VCFTOOLS to obtain pairwise relatedness statistics between all individuals (Yang et al., 2011) prior to downstream analyses. Individuals that are very closely related have the potential to skew patterns of population structuring. The expectation for this function is that comparisons of theoretically unrelated individuals within populations are assigned a value of 0 and individuals compared against themselves have a value of 1.

## 2.4 | Characterizing patterns of genomic divergence in the San Francisco Bay

We employed several programs to identify patterns of genetic substructure while investigating patterns of diversification among sparrow populations around the bay region. We used Populations

in STACKS to measure variation across the 12 populations representing the six subspecies by calculating pairwise  $F_{ST}$  and the observed and expected heterozygosity. Using the R package "POPPR" (Kamvar et al., 2015), we analysed hierarchical genetic structuring using analysis of molecular variance (AMOVA). We conducted 999 permutations to test for significance in  $F_{IT}$ ,  $F_{IS}$ , and  $F_{ST}$ .

To visualize genetic clustering among subspecies, we conducted principal component analysis (PCA) using the R package "SNPRELATE" (Zheng et al., 2012). We analysed patterns of genetic structure in a Bayesian framework using the program STRUCTURE version 2.3.4 (Pritchard et al., 2000). We conducted 10 runs for each value of K = 1-7 for our six subspecies; each run consisted of a 200,000 iteration burnin followed by 300,000 sampling iterations. We implemented the admixture model and used an allele frequency prior that was estimated from an initial run of K = 1 ( $\lambda = 0.29$ ). We determined the most likely number of clusters (K) using the method described by Evanno et al. (2005) as implemented in STRUCTUREHARVESTER version 0.6.94 (Earl & vonHoldt, 2012). We averaged results across all of our runs using the greedy algorithm in CLUMPP version 1.1.2 (Jakobsson & Rosenberg, 2007) and visualized results using DISTRUCT version 1.1 (Rosenberg, 2004).

# 2.5 | Spatial patterns and the influence of geographic & environmental isolation

We evaluated the role of spatial patterns, specifically geographic isolation, in influencing genetic structure among populations in the bay region. We first used the R package "CONSTRUCT" (Bradburd et al., 2018) to visualize continuous and discrete population structure using both spatial and nonspatial models. CONSTRUCT differs from traditional model-based clustering methods designed to detect discrete population structure (i.e., STRUCTURE) in that it jointly models the effects of both discrete population structure and continuous isolation by distance on sample relationships (Bradburd et al., 2018). We ran CONSTRUCT for K = 1-7 both with and without spatial information, with 10,000 iterations for three independent chains, and subsequently compared these models using cross-validation analysis. If the predictive accuracy of the spatial model is higher than the nonspatial model, it is assumed that spatial patterns (i.e., isolation by distance) are contributing to the observed genetic structure. We also tested for isolation by distance (IBD) using the R package "ADEGENET" (Jombart & Ahmed, 2011), using locations from all 12 sampling sites.

We additionally visualized spatial population structure and genetic dissimilarity between populations using estimated effective migration surfaces (EEMS; Petkova et al., 2016). EEMS models the genetic and geographic relationships using matrices of average pairwise genetic distance and geographic distances between individuals. The program models effective migration based off of the stepping stone model and Markov Chain Monte Carlo sampling from the posterior probability distribution of the genetic dissimilarities. We calculated pairwise genetic distance using the "bed2diffs" function MOLECULAR ECOLOGY – WILEY

in EEMS and geographically referenced all individuals using respective sampling site locations. We ran EEMS using three independent chains each with 10,000,000 MCMC iterations with a 20,000,000 burnin and a population grid density of 700 demes, adjusting all other parameters until achieving a proposal acceptance rate between 20% and 30%. We used the R package "rEEMSplots" to plot model fits (to confirm chain convergence for each individual run), and to generate contour maps of effective migration and nucleotide diversity.

We evaluated whether the magnitude of genetic differentiation  $(F_{ST})$  between the salt marsh populations and the upland populations could be attributed exclusively to genetic drift. To do this, we used linear regression analyses to correlate pairwise estimates of  $F_{ST}$  between each salt marsh population versus *gouldii* (chosen as a representative upland population) as a response variable and various measures of genetic diversity as the predictor variables (Funk et al., 2016). For the predictor variables, we used estimates of observed and expected heterozygosity (estimated in STACKS using only variant sites) and nucleotide diversity (estimated in STACKS using all positions, variant and fixed). Here, we assume that if genetic drift is responsible for driving high levels of genetic differentiation, we should see a negative correlation between population diversity and pairwise  $F_{ST}$  between marsh and upland populations.

Lastly, we evaluated whether the magnitude of genetic differentiation ( $F_{ST}$ ) could be attributed to local differences in environment. We performed partial-mantel tests using the R package "vEGAN" (Oksanen et al., 2019) to test for evidence of adaptive differentiation between subspecies considering environmental factors, in addition to pairwise  $F_{ST}$  and geographic distance. We obtained environmental data on water salinity and air temperatures of different marshes and upland habitats from USGS and NOAA repositories (NOAA, 2020; Schraga & Cloern, 2017).

### 2.6 | Identifying putative outliers

To assess whether selection is contributing to the fine scale diversification in the bay region, we scanned for outliers using two different approaches. For both of these approaches, we used the full data set (10,270 SNPs). First, we identified outlier SNPs using BAYESCAN version 2.1 (Foll & Gaggiotti, 2008). We scanned all SNPs in pairwise comparisons across all of our subspecies and allowed for a false discovery rate of 5%. Second, we used per locus SNP  $F_{ST}$ estimates (calculated in VCFTOOLS; Danecek et al., 2011) to identify putative outliers in our data set. For each pairwise comparison, we characterized a SNP as having elevated levels of differentiation if it had an  $F_{ST}$  estimate greater than five standard deviations above the mean. To assess the contribution of neutral versus potentially adaptive SNPs in delineating population boundaries, we repeated the PCA using only the SNPs identified as putatively under selection using either method. For the F<sub>ST</sub> approach, a SNP had to be classified as elevated in more than one pairwise comparison. For all outlier SNPs, we assessed the homology and the distribution of loci across the Zebra finch (Taeniopygia guttata) genome using BLASTn (Altschul

gouldii	heermanni	santaecrucis	maxillaris	pusillula
0.0178				
0.0168	0.0156			
0.0157	0.0136	0.0137		
0.0235	0.0224	0.0197	0.0190	
0.0181	0.0169	0.0166	0.0138	0.0232
	gouldii 0.0178 0.0168 0.0157 0.0235 0.0181	gouldii         heermanni           0.0178	gouldii         heermanni         santaecrucis           0.0178         -         -           0.0178         0.0156         -           0.0168         0.0136         0.0137           0.0157         0.0136         0.0137           0.0235         0.0224         0.0197           0.0181         0.0169         0.0166	gouldii         heermanni         santaecrucis         maxillaris           0.0178         -

et al., 1990). To assign matches, we required an E value of  $<\!1.0\times10^{-}$   $^{5}$  and  $>\!70\%$  identity score.

#### **TABLE 2** Pairwise $F_{ST}$ estimates for the six subspecies of song sparrow in San Francisco Bay. The average $F_{ST}$ across all subspecies is 0.018. *M. m. pusillula* accounts for the most variation, and *M. m. santaecrucis* the least

#### 3 | RESULTS

#### 2.7 | Demographic models

To estimate the demographic history of song sparrows in the bay region, we fit a series of demographic models to the site frequency spectrum using the program moments (Jouganous et al., 2017). For input, we used a VCF file with no missing data that included 4,366 SNPs sampled from across 82 individuals: 13 M. m. gouldii, 17 M. m. heermanni, 15 M. m. maxillaris, 20 M. m. pusillula, and 17 M. m. samuelis. The VCF was converted to a moments input format using a custom python script from Isaac Overcast (https://github.com/ isaacovercast/WCS/blob/master/vcf2dadi/vcf2dadi.py). We first inferred divergence times, population sizes, and gene flow following an initial split between M. m. pusillula and all other subspecies. For all demographic models, we compared a model with gene flow following population divergence to one without. We also compared a history of constant population size, exponential growth, or a bottleneck in either M. m. pusillula, the other subspecies, or in both populations (see Figure S1 for illustration of all models compared). We additionally estimated divergence time and migration history among three populations (M. m. pusillula, M. m. gouldii, and all others).

For all two-population models, 15 optimizations with maximum iterations of 30 were run from different starting parameters using the perturb function in moments. We then ran one final optimization with 75 iterations using the parameter values estimated from the shorter run with the highest likelihood. To increase computational speed for three population models, we ran 10 optimizations with maximum iterations of 10, and a final run with 50 iterations. We calculated demographic parameter values from the estimated value of theta (4N $_{e}\mu$ L; L is sequence length) based on the assumption of a one year generation time for song sparrows (Arcese et al., 2002), the average substitution rate for Passeriformes  $(3.3 \times 10^{-9} \text{ substitutions/site/year; Zhang et al., 2014})$ , and total sequence length equal to 260,662 bp (2,773 loci  $\times$  94 bp/loci). Parameter uncertainties were calculated using the Godambe information matrix (GIM; Coffman et al., 2016). Finally, to infer the best fit model, we performed a likelihood ratio test with a GIMbased adjustment to account for the use of composite likelihoods (Coffman et al., 2016).

We ran the STACKS pipeline on a total of 157 individuals (totaling 300,342 reads). Our samples had an average alignment rate to the reference genome of 95.2% and an average depth of coverage of 35.7X. We removed four samples with more than 50% missing data and one sample due to 83% relatedness to another individual, resulting in a final sample size of 152 sparrows.

#### 3.1 | Genomic divergence in the San Francisco Bay

PCA revealed a modest signal of genetic variation among song sparrow populations, with M. m. pusillula demonstrating the clearest signal of divergence from other subspecies (Figure 1b). PC1 and PC2 explained 1.47% and 1.25% of the variation, respectively. While subsequent axes explained similarly low amounts of variation, PC2 and PC3 suggest a pattern of differentiation between the upland subspecies, gouldii and heermanni from those restricted to the Bay (Figure 1b; Figure S2). AMOVA results revealed that variation among individuals ( $F_{1S}$ ; p = .082, 1.877%) and among subspecies ( $F_{ST}$ ; p = .001, 1.984%) was lower than expected, but that variation within individuals ( $F_{IT}$ ; p = .003, 96.139%) was greater than expected. Only *p*-values from  $F_{ST}$  and  $F_{IT}$  are statistically significant. Average observed heterozygosity was similar across all of the subspecies with the exception of pusillula, which was slightly lower (Table 1). Pairwise  $F_{\rm ST}$  estimates supported the results of the PCA (Table 2). The global  $F_{sT}$  is 0.018 (0.0136 <  $\mu$  < 0.0235), which represents a moderate amount of differentiation given the small spatial scale. The highest differentiation occurs between pusillula and all other subspecies  $(0.0196 < \mu < 0.0235)$  and the lowest differentiation is between san*taecrucis* and all other subspecies (0.0136 <  $\mu$  < 0.0196). Pairwise  $F_{sT}$  estimates further suggest increased differentiation among the three marsh subspecies (maxillaris, pusillula, samuelis) than within their upland counterparts. No pairwise comparisons were statistically significant.

Results from STRUCTURE were consistent with our PCA, with *pusillula* separated from the other populations most strongly. With all six subspecies included, the  $\Delta K$  method in STRUCTURE identified K = 2 to be the optimal number of clusters, reflecting a clear split of *pusillula* from other subspecies (Figure 2). We subsequently performed a STRUCTURE run without *pusillula* to see if there was additional structuring being obscured. We estimated a new allele frequency prior

 $(\lambda = 0.31)$  for the five subspecies included in this latter run, and the  $\Delta K$  method supported K = 3 as the optimal number of clusters (Figure 2). Based on Q values (an individual is assigned to a cluster based on majority percent cluster assignment), hierarchical STRUCTURE results identified one defined cluster comprised of *samuelis* individuals (80% of *samuelis* individuals were assigned to this cluster). STRUCTURE also identified a second, moderately less defined cluster, that corresponded to *gouldii* individuals (54% of *gouldii* individuals were assigned to this cluster). The third cluster was comprised of individuals from the three remaining subspecies (*maxillaris, heermanni*, and *santaecrucis*; Figure 2), which appear to be largely undifferentiated from each other. We note here that while it was not the best supported model, K = 5 does suggest some additional substructuring among *maxillaris, heermanni*, and *santaecrucis* (Figure S3).

# 3.2 | Spatial patterns and the influence of geographic and environmental isolation

Cross-validation analysis of the CONSTRUCT results revealed no significant difference between the spatial and nonspatial models, indicating that isolation by distance is not responsible for the differentiation we are observing (Figure S4). We found no significant correlation between genetic and geographic distance among the song sparrow populations in the bay region (p = .995, Figure S5).

Effective migration rates, estimated using EEMS, are low into *maxillaris* and *pusillula* populations, and nucleotide diversity rates are notably low within *samuelis* and *pusillula*. The effective migration rates are visualized and plotted on a log<sub>10</sub> scale as a contour map (Figure 3) with colours representing estimated (a) average migration rates, m; and (b) average nucleotide diversity surfaces, q, ranging from low (dark red) to high (dark blue; Figure 3). There are apparently no strict barriers to migration, but we observe notably lower migration rates and genetic diversity for each of the three populations restricted to single arms of the Bay.

We did not observe clear evidence of genetic drift through comparisons of pairwise  $F_{ST}$  between each saltmarsh population and gouldii (as a representative upland group) and measures of genetic diversity (Figure S6;  $H_0$ :  $R^2 = -.118$ , p = .572;  $H_E$ :  $R^2 = .266$ , p = .135;  $\pi$ :  $R^2 = -.1654$ , p = .716). However, we did observe a notable positive trend in a partial-mantel test correlating pairwise  $F_{ST}$  and salinity (while controlling for distance; p = .07; Figure S7). There was no significant effect of temperature on pairwise  $F_{ST}$  (p = .5).

### 3.3 | Identifying putative outliers

Based on two approaches for outlier detection, we detected several candidate SNPs that are putatively under selection. BAYESCAN identified two outliers from the full data set (0.019% of all SNPs; Figure S8). The two identified outlier SNPs had  $F_{ST}$  values of 0.11193 and 0.11335. Using a  $F_{ST}$  cutoff approach, we identified a number of SNPs that exhibited elevated  $F_{ST}$  estimates (functionally defined here as five standard deviations above the mean), ranging from 19 (gouldii-pusillula) to 37 (maxillaris-pusillula) putative outliers across the fifteen pairwise comparisons. Of these putative outliers, 103 elevated sites were shared across more than one pairwise comparison. PCA using only these 103 shared SNPs resulted in a similar pattern as the unlinked data set, with a slightly more defined separation between salt marsh and upland populations (Figure S9). Several of these putative outlier regions aligned to annotated regions of the Zebra Finch genome (Table S2).

#### 3.4 | Demographic history

An isolation with continuous migration (IM) model was the best fitting model to the joint site frequency spectrum of *pusillula* and all other subspecies (Figure 4; log-likelihood: -608.19). An IM model with exponential growth in the other subspecies population (LL: -607.6) and an IM model with a bottleneck within the *pusillula* population (LL: -607.93) both showed slightly greater likelihoods relative to the IM constant size model. However, likelihood ratio tests comparing the IM constant to the growth model (adjusted D: 0.0139;



**FIGURE 2** Results from the STRUCTURE analysis. For all subspecies, K = 2 is most optimal, separating *pusillula* from the rest of the subspecies. A hierarchical run excluding *pusillula* suggests K = 3 as the best, separating the marsh endemic *samuelis* and the upland *gouldii* 

#### (a) Posterior mean migration rates



(b) Posterior mean diversity rates



**FIGURE 3** Estimating effective migration surfaces (EEMS) contour plots for effective migration (a) and nucleotide diversity (b). Average migration and diversity rates are represented as a colour gradient ranging from low (orange) to high (blue). Effective migration rates are low among *maxillaris* and *pusillula* populations, and diversity rates are notably low among *samuelis* and *pusillula*. There is less gene flow and genetic diversity in the subspecies endemic to the San Francisco Bay Area

p = .453) and IM constant versus. IM bottleneck (adjusted D: 0.535; p = .232) both show that increasing model complexity does not result in a significantly better fit to the site frequency spectrum. Under the IM constant population size model we inferred that pusillula diverged from the other subspecies ~61 kya (95% CI: 23,560-113,500 ya). Following the split *pusillula* maintained an effective population size of ~26,573 (95% CI:10,119-49,622 ya) and the other four subspecies an N<sub>o</sub> of 256,940 (95% CI: 168,023-362,041 ya). Migration was found to be slightly greater from other subspecies into pusillula  $(1.37 \times 10^{-4})$  than in the opposite direction  $(2.82 \times 10^{-5})$ . Uncertainties surrounding divergence time between pusillula and the other subspecies were large with standard deviations inferred using the GIM method exceeding the parameter estimate (Table S3). This wide distribution reflects a bimodal distribution of divergence times with most divergence time estimates less than 100 kya, but a second cluster showing divergence times >250 kya (Figure S10). The threepopulation model (log-likelihood: -1659.93; Figure S11) showed a much deeper divergence between pusillula and the other subspecies

at ~527,190 generations ago (95% CI: 467,215–601,846) followed by divergence between *gouldii* and all other subspecies more recently, ~2,156 generations ago (95% CI: 1–16,146). Effective population size estimates were larger for *pusillula* (50,984; 95% CI: 39,512–63,725), but there was overlap in confidence intervals between two-populations and three-population analyses. *Gouldii* exhibited the largest effective population size (233,035; 95% CI: 210,983–255,739), while the remaining three subspecies combined had an estimated N<sub>e</sub> comparable to *pusillula* (53,102; 95% CI: 41,783–65,637). Migration rate was also found to be highest between *gouldii* and *pusillula* (3.01 × 10<sup>-5</sup>; 95% CI: 0–9.72E-05), though confidence intervals did encompass zero, suggesting the possibility of no gene flow between these subspecies.

## 4 | DISCUSSION

We describe patterns of genetic structure in San Francisco Bay song sparrows to elucidate forces capable of driving species diversification in the face of ongoing gene flow. Within a  $70 \times 100$  km region, we identified modest signals of differentiation of the subspecies pusillula, samuelis, and gouldii. We note here that although the remaining subspecies in the Bay (heermanni, santaecrucis, maxillaris) appeared largely undifferentiated from each other in the best supported STRUCTURE model, we observe some continued substructuring at greater values of K. Further, despite hypotheses that santaecrucis is a hybrid between gouldii and heermanni, STRUCTURE results suggest signals of ancestry (either linked to gene flow or shared ancestral variation) from samuelis as well-although future studies that combine increased sampling efforts along a transect would be more appropriate to address this in full. While the observed differentiation in all cases was subtle, the patterns observed here are notable given the fine spatial scale over which this study was conducted. Moreover, signals of genomic differentiation between tidal marsh and upland clades coupled with evidence of isolation by environment support earlier suggestions that natural selection has played at least a partial role in driving this differentiation (e.g., Basham et al., 1987; Chan & Arcese, 2003; Johnston, 1956b; Miller, 1956). We estimated that the present-day diversification of song sparrows in the bay region occurred recently (within the last 500,000 generations). While the split between *pusillula* and other subspecies predates marsh formation, estimates for the other groups are more consistent with the recent formation of salt marsh habitats within the bay region. However, the divergence times estimated here were not well-resolved (see below). Overall, our results offer new insight into the roles of natural selection and demography in shaping divergence of song sparrow populations in this region.

Several mechanisms are proposed to explain the diversification of song sparrows in the bay region. Aldrich (1984) suggested that ecological gradients including salinity, climate, seasonality, and background coloration have all contributed to microevolutionary change in song sparrow across their range, but found little support for historical effects of geographic isolation or genetic drift on phenotype.



FIGURE 4 (Left-hand panel) Schematic of best-supported model, where M. m. pusillula splits from all other subspecies ~61 kya. Following divergence continuous gene flow continued between both populations with slightly greater migration into M. m. pusillula. The four panels on the right side of the plot show how well the best-fit model fits the observed site frequency spectrum. The top-left panel shows the observed SFS. The top-right panel shows the simulated SFS based on the best-fit model and parameters. The bottom panels show the residuals from fitting observed data to model data

Patten & Pruett (2009) described song sparrows as a ring species due to the pattern of subspecific morphological variation over their geographic range. Our results provide further insights into the origins of rapid differentiation with the San Francisco Bay region, specifically with respect to the roles of three potential drivers of divergence: isolation, drift, and natural selection. First, our analyses using CONSTRUCT failed to explain observed patterns better than non-spatial models, offering no evidence of isolation by distance among these sparrow populations. Although geographic isolation has been proposed as a mechanism shaping diversification in song sparrows within and outside the bay region (e.g., Ferrell, 1966; Miller, 1947; Patten & Pruett, 2009), the spatial scale over which isolation by distance occurs should be proportional to the dispersal distance of an organism and the extent of gene flow between populations. Thus, this scenario seems less likely given that we expect the dispersal capabilities of these fully flight-capable sparrows to far exceed the spatial scale over which we found genetic divergence.

Evidence for a role of drift in promoting population divergence among subspecies would include low genetic diversity, small effective population sizes, and/or a history of population bottlenecks. EEMS analyses show lower nucleotide diversity within tidal marsh populations of song sparrow (Figure 3). While analysis of the site frequency spectrum did not support a history of bottlenecks in these populations, previous analyses using microsatellite loci from many of the same individuals (Chan & Arcese, 2002) and whole genome sequencing (WGS) data from M. m. pusillula and M. m. gouldii (Walsh et al., 2019a) reveal contrasting results. Specifically, WGS analysis found evidence for recent bottlenecks (<250 years ago) in both M. m. pusillula and M. m. gouldii, and much smaller current N<sub>2</sub>: 324 and 179, respectively. It is likely that there is more power in the larger

WGS data set to detect fluctuations in N<sub>a</sub> (e.g., Terhorst et al., 2017), as well as to detect more recent fluctuations in population size compared to the RAD data. Additionally, the difference in population sampling between our studies could contribute to different results. Gene flow with unsampled populations can lead to erroneous inferences of population bottlenecks (e.g., Nielsen & Beaumont, 2009) as well as bias estimation of effective population size (e.g., Lynch & Sethuraman, 2019). While further analyses will be required to resolve these differences in demographic history among different data sets, the inferred N<sub>o</sub> and patterns of genetic diversity across all data sets exceed those observed in other species where drift appears to have played a significant role in population differentiation. For example, Funk et al. (2016) found that in island fox (Urocyon littoralis) populations, Ne was inferred to be as low as 2.1 (range: 2.1-89.7) and heterozygosity as low as 0.016 (range: 0.016-0.231). Despite the potentially strong influence of drift for those fox populations, the authors still detected signatures of selection in comparison of their genomes across island populations. In sum, our results and prior work on song sparrows from the San Francisco bay region provide mixed support for drift driving population differentiation. While it is probable that drift contributed to the observed patterns to an extent, it is highly unlikely that drift alone shaped patterns and levels of population differentiation among these subspecies.

We found support for selection in driving diversification of sparrow populations around the Bay, possibly related to adaptation to saltwater versus freshwater habitats. The hypothesized role of selection in shaping patterns of differentiation is not unreasonable given the steep ecological gradients and adaptive challenges associated with freshwater-saline ecotones (Greenberg et al., 2006; Walsh et al., 2019a). These patterns are notably different, however, from WII FY-MOLECULAR ECOLOGY

comparisons of tidal marsh Savannah Sparrows in San Francisco Bay, which exhibited low levels of genetic differentiation, even across ecological gradients (Benham & Cheviron, 2019). Pairwise comparisons resulted in the identification of 103 shared SNPs that exhibited elevated signals of differentiation ( $F_{ST}$  greater than 5 SD above the mean). Some of these differentiated regions (Table S2) corresponded to genes with putative functions in osmoregulatory processes, including genes associated with ion homeostasis (Di Ciano-Oliveira et al., 2006), regulation of MAPK cascades (Kültz & Avila, 2001; Vom Dahl et al., 2001), and microtubule cytoskeleton organization (Di Ciano-Oliveira et al., 2006). Of particular interest are several genes with putative links to cytoskeleton organization (MYH13, MYO18, MYO3, TTBK2, PAK3; Table S2) that are associated with signals of elevated divergence between upland and salt marsh populations of San Francisco Bay song sparrows. Previous work has shown that the cvtoskeleton, which is responsible for maintaining cell morphology. is a likely candidate for regulating cell volume response in the face of osmotic changes (Di Ciano-Oliveira et al., 2006; Bober et al., 2015). Cytoskeletal reorganization may allow cells to resist volume changes through the reinforcement of cell structure (Di Ciano-Oliveira et al., 2006), a potentially important adaptation to salt water environments. Putative selection for genes linked to cytoskeletal organization have been identified in other freshwater-saline comparisons, including in savannah sparrows (Walsh et al., 2019a) and saltmarsh sparrows (Walsh et al., 2019b). Savannah sparrows (Benham & Cheviron, 2020; Goldstein, 2006) and Chilean seaside cinclodes (Sabat et al., 2004) both show elevated plasma osmolality in high salinity environments, suggesting that genes related to cytoskeletal reorganization may be important for increased salinity tolerance in these populations. These findings paired with whole-genome comparisons of M. m. gouldii and M. m. pusillula, which identified similar candidate genes with putative links to tidal marsh adaptations (Walsh et al., 2019a) support a role for microgeographic adaptation in shaping patterns of divergence in this region.

Independent of their mechanism of origin, these newly characterized fine scale patterns of diversification among song sparrow populations in the San Francisco Bay region have tangible conservation and management implications. The loss of marsh habitat threatens the breeding and nesting grounds for all three Bay-endemic subspecies (maxillaris, pusillula, and samuelis), which are the only extant populations of song sparrow adapted to life in salt marshes. It has been estimated that the current marsh habitat occupies only 15% of its pre-industrial acreage (Marshall & Dedrick, 1994), and projected sea level rise poses serious threats to the persistence of these intertidal ecosystems. Based on current population declines, updated information on local adaptive capacity and potential management units within the Bay has significant conservation relevance. Our research provides evidence for genetic structuring on a fine spatial scale, with a putative role of local adaptation in driving population diversification. A major challenge facing conservation and evolutionary biologists is the identification and preservation of biological diversity and ecological viability in response to anthropogenic change. Characterizing and maximizing genetic variation should be a

key goal in any management endeavor, as increased genetic diversity can enhance a population's resilience in the face of changing environmental conditions (Reed & Frankham, 2003). The historical designation of five subspecies within San Francisco Bay Song Sparrows is supported by our results and, we hope, this information can direct meaningful conservation efforts in the region. While the utility and applicability of subspecies as units of conservation and taxonomy remains a topic of debate, our work suggests that forces of local selection are counteracting gene flow between these subspecies, supporting the potential for adaptive genetic diversity on a population-scale.

Based on our findings, we recommend that local management initiatives strongly consider the potential for locally adapted populations (on the scale of subspecies) within the bay region, with a particular focus on the *pusillula* subspecies given the clear differentiation of this group from the remainder of the populations in the region. More generally, these factors may be particularly important if considering any future translocation or captive breeding efforts.

In conclusion, our results offer evidence of local adaptation at a microgeographic scale in subspecies of song sparrows residing in the marshes and uplands surrounding San Francisco Bay, including signals of differentiation in two of the three salt marsh-dwelling forms. Although correlative with regard to mechanism, our results offer support for the hypothesis that context-dependent natural selection on individual performance has played a role in the diversification of song sparrows in this region; and specifically, that local adaptation to salt marsh environments and genetic drift have each contributed to the variation in population structure we describe. Although differentiation across subspecies was modest overall, a putative role for natural selection in shaping these patterns was suggested by whole-genome analyses of two of the subspecies studied here (Walsh et al., 2019a). We propose that whole-genome comparisons of song sparrows targeting loci or genomic pathways under selection, and thus putatively linked to underlying local adaptation, are now warranted. Incorporating such information into scenario planning and management decisions in the San Francisco Bay region offers further opportunities to maximize the persistence of this and similarly variable species by developing plans most likely to conserve the adaptive scope of organisms at their appropriate geographic scales.

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#### AUTHOR CONTRIBUTIONS

This study is part of the undergraduate honors research of C.M under the supervision of J.W. and I.J.L. C.M., I.J.L., and J.W. conceived and designed the study, with input from P.A. and Y.C. Y.C. and P.A. collected samples. C.M. and J.W. conducted laboratory work. C.M., S.M.A., J.W., and P.B. analysed the data. S.M.A., C.M., and P.B. prepared and created figures for the manuscript. C.M. and J.W. wrote the manuscript based on input and comments from all authors, especially I.J.L. and P.A. All authors read and approved the final manuscript.

#### DATA AVAILABILITY STATEMENT

Filtered VCFs are deposited in Dryad: https://doi.org/10.5061/ dryad.ncjsxkssb. Any additional data that support the findings of this study are available from the corresponding author upon reasonable request.

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#### SUPPORTING INFORMATION

Additional supporting information may be found online in the Supporting Information section.

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